

CELL CULTURE TECHNICAL PROCEDURES

- XX. 100 Cell Lines and Preparation for Test
- XX. 101 Lines: Use P388 **or** L1210 cells.
- XX. 102 Stock Cultures: Cultivate in **a** stationary flask (cells will not adhere) in Fischer's Medium plus 10% horse serum. Twenty-four hours before test prepare **a** spinner flask culture to obtain **cells** in logarithmic phase of growth.
- XX. 103 Preparation of Cell Suspension: Dilute spinner cell suspension to 6.6×10^4 cells per ml (this will be 5×10^4 cells per ml in final test mixture). Deliver 1 ml of test material and 3 ml of cell suspension to culture tubes. Incubate at 37°C for 48 hours. The baseline count (Co) will be 5×10^4 .
- XX. 104 Preparation of Materials: See 13.104.
- XX. 200 Dose Levels: All synthetics and plant products are to be tested by weight (W), not dilution. First tests **of** synthetics and **all** priority plant fractions are **to** be scheduled at 100, 10, 1, 0.1, and 0.01 ug/ml . All plant crudes and B002 samples are tested at 100, 10, and 1 ug/ml . Fermentation products with **a** sample code of D through K may be tested by weight **or** dilution. All other fermentation products must be consistent (W or D) within the NSC core number. When tested by dilution, starting doses are 1:10, 1:100, and 1:1000 (one point pre-screens are done at **a** dilution of 1:50).
- XX.300 Experimental Design: Use a common set of control tubes to evaluate materials tested at one time (about 40). Use the number of control tubes according **to** the formula $2\sqrt{n}$ where n = number of materials being tested.
- XX.400 Calculations: Cell counts are made for all test and control tubes by an appropriate means: hemocytometer, Coulter counter, etc. For an automatic counter, it may be necessary to derive a calibration factor by comparison **of a** hemocytometer count of control cells with the machine count. Duplicate counts of each tube may be desirable. Routinely, a 1:20 dilution of control and test tubes is made in a medium appropriate to the cell counter being used.
- XX.401 The mean value of all control tubes will be used for calculations of C.
- XX.402 The mean of the experimental tubes (T) for each dose (dilution) minus the mean of the baseline (Co) divided by the mean of the control tubes (C) minus Co **give the** growth ratio (Y) at each dose level. Multiply by 100 and express as percent.

$$100 \times \frac{T - C_0}{C - C_0} = Y \%$$

XX.403 The slope is the difference in response for a one-log difference in **dose**, calculated by linear least squares regression, as follows:

- A. If the growth ratios (i.e., Y values) computed for each dose are all greater than 55%, do not compute the slope. Indicate that the ED50 is greater than the maximum dose for weight formulation, or less dilute than the smallest dose for testing by dilution.
- B. If the Y values are all less than 45%, bypass the slope calculation and indicate that the ED50 is less than the minimum dose (weights), or more dilute than the greatest dose (for dilution testing).
- C. Otherwise, compute the slope and intercept of the regression line as follows. Note -- do not use more than one point from the region $Y \leq 15\%$; similarly, only one point from the region $Y \geq 85\%$ should be used.

N = number of points selected. $\{\leq \text{number of dose levels} \ \& \ \geq 2\}$
 X_i = \log_{10} of dose_i
 Y_i = growth ratio calculated for dose_i
 Y = $A + B \cdot X$ regression line

$$B = \text{slope} = \frac{N \cdot \sum(X_i \cdot Y_i) - (\sum X_i) \cdot (\sum Y_i)}{N \cdot \sum(X_i)^2 - (\sum X_i)^2}$$

$$A = \text{intercept} = \frac{\sum Y_i}{N} - B \cdot \frac{\sum X_i}{N}$$

XX.404 The ED50 is the effective dose **which inhibits growth to 50% of control** growth, calculated as follows:

- A. Using the slope and intercept from the regression line, compute the log of the effective dose which inhibits growth to 50% of control **growth**:

$$\begin{aligned}
 Y &= A + B \cdot X \text{ regression line} \\
 50\% &= A + B \cdot (\log \text{ED50})
 \end{aligned}$$

$$\log \text{ED50} = (50-A) / B$$

B. For materials tested by weight, the ED50 dose is the antilog of the value computed above, i.e.:

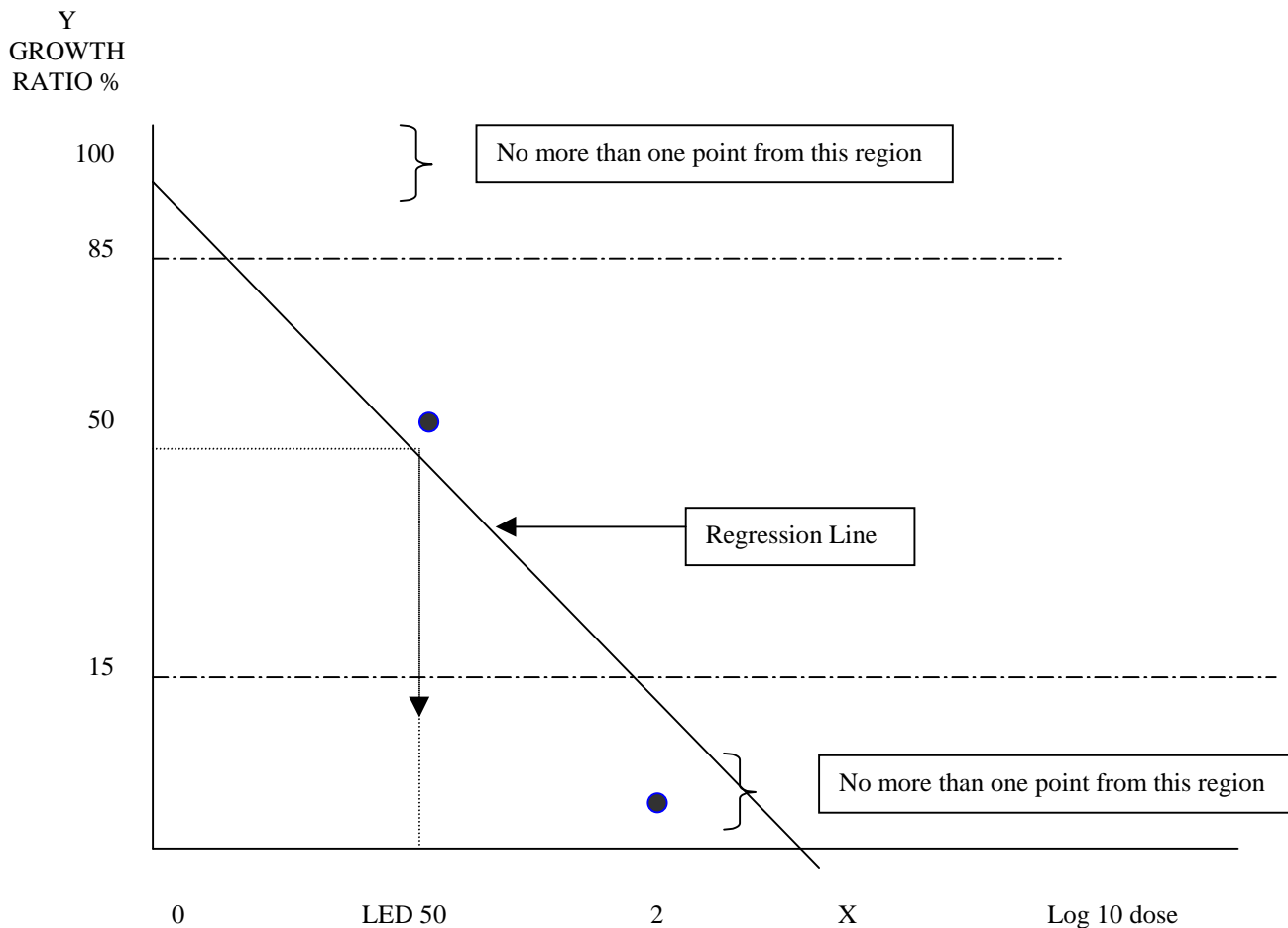
$$\text{ED50} = 10^{(\log \text{ED50})} \text{ ug/ml}$$

C. For materials tested by dilution, the ED50 dilution is the antilog of the negative log ED50, i.e.:

$$\text{ED50} = 10^{(-\text{LOG ED50})} \text{ dilution}$$

EXAMPLE;

If $\log \text{ED50} = -3.0$,
then $\text{ED } 50 = 1000$ dilution
which represents a concentration of 1:1000



XX.500 Quality Control

- A. The 48-hour control tubes shall show growth of at least ten times that of the baseline values.
- B. The maximum allowable difference between cell counts of duplicate tubes at each dose level has not been set.
- C. The limit for reversal of Y values between consecutive dose levels has not been set.
- D. The positive control compound, NSC 95441 (MeCCNU), is tested in every experiment. Quality control limits for MeCCNU are tentatively set at ED50=1.7-7.7 ug/ml.

XX.600 Test Evaluation: These parameters **have** not been set.

XX.700 Number and Interval of Dose Levels: See 13.700

XX.800 Priorities: These are as set by the DR&DP.